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Environmental radiation of protozoa



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Abstract

The evaluation of some of the findings on the impact of UV-B radiation on ciliated protozoa in the literature focuses in particular on the changes in motility and photomotility, two factors that are crucial in determining an organism's capacity to survive in its environment. In a moment, it will be explained what ciliates are and why they are crucial to ecological systems. The early studies that looked at how UV light affected ciliates will be summarised here. The findings of research on Fabreasalina, a marine ciliate, and Blepharismajaponicum and Ophryoglenaflava, two freshwater ciliates, will next be discussed in greater depth.

Keywords: UV Radiation, motility and photomotility, Fabreasalina

1. Introduction

Environmental science is often referred to in relation to nature. The vastness of nature is home to a diverse array of interconnecting levels of connection, ranging from minute-scale (such as cells) to planetary-scale (such as biosphere) anomalies. Regarding how a certain aspect of the environment, such as light, water, and nutrients, influences a living cell, a specific organ, the complete individual, or a group of people, several perspectives may be taken on how organisms relate to their present situation. The four degrees of connection that biology is primarily concerned with are population, local area, environment, and bio frameworks. Environment is not a static system made up of abiotic and biotic components. Instead, it is a dynamic, self-adjusting structure in which biotic and abiotic elements interact.

1.1.Anaerobic degradation of organic compounds

Anaerobic corruption of natural mixtures the responses in an anaerobic digester are complicated with various successive and equal advances and can be separated into two principal types

(a) Biochemical responses: These are regularly catalyzed by intra or extracellular catalysts and follow up on the pool of naturally accessible natural material. Crumbling of composites (like dead biomass) to particulate constituents and the resulting enzymatic hydrolysis of these to their solvent monomers are primarily extracellular. Corruption of solvent materials are interceded by life forms intracellularly, bringing about biomass development and resulting rot.



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(b) Physico-synthetic responses: These are not naturally interceded and include particle affiliation/separation, fluid strong and gas-fluid exchange.

2. Review of Literature

Water performs several essential roles in habitats all around the planet Earth. It is one of our most important regular resources since life cannot exist without it. Understanding the aquatic ecosystem is essential since water covers a large portion of the planet. Amphibian biological systems have a variety of important environmental activities, including as recycling nutrients, purifying water, reducing floods, reviving ground water, and providing habitat for wild animals. It is also used for human amusement and is essential to the operations of the fishing and tourist industries. Early sources of knowledge on freshwater life, comparable to those about marine life, appeared in the distant past, maybe before the time of Aristotle (384-322 BC). These early findings, which usually include bizarre concoctions of the real and the spectacular, are illogical (Welch, 1952). With the passage of time and the progressive expansion of man's knowledge of his surroundings, he started learning more about each component of nature. Numerous scholars are interested in limnology, the field that oversees the exploration of all inland oceanic environments, including streams, rivers, lakes, supplies, and wetlands. The public was made aware of Francois Alphonse Forel's (1841–1912) efforts via his early research in Lac Leman (the Geneva, Switzerland, pool). The topography, physical science, and lake science are covered in the first two volumes, which were published in 1892 and 1895, respectively. The lake science is covered in the third book, which was published in 1904. Earlier, in 1896, Forel published and disseminated a report on the Lake Geneva's base fauna. He delivered the essential limnology readings in 1901. At that time, Forel, a professor at the College of Lausanne, was perhaps recognised as the "Father of Limnology."

Throughout the first half of the 20th century, limnology continued to develop as a topic of study and expanded its geographic scope. In the 1920s and 1930s, limnologists created a large number of field stations, made extensive use of them to collect copious amounts of data on specific lakes, and organised this data at the provincial level. The study of moderate limnology began the foundation of knowledge on the physical, chemical, and ecological characteristics of lakes, which was completed by Birge and Juday (Wetzel, 1996). It is important to recognise a



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few individuals who have achieved success in the field of limnology around the globe. Other pioneering workers besides Forel include Stephen Alfred Forbes (1844-1930).

3. Methods

Lashes:- During 2010–2011, the total number of whips per millilitre was 6780 at Station 1, 4600 at Station 2, and 656 at Station 3. At stations 1, 2, and 3 independently, the numbers were 6670, 4984, and 490 in the 2011–12 academic year. The results of the population's station-wise analysis revealed that for both years, station 1 had the thickest population and station 3 had the thinnest. At each site, the beat population density increased during the winters, decreased during the rainstorms, and decreased during the summers of both years (Table 1).

Station 1:

2010-11

| Sr.No. | Flagellates | Monsoon | Winter | Summer | Total |
|--------|--------------------|---------|--------|--------|-------|
| 1. | Zoomastigophorea | 1402 | 1520 | 964 | 3886 |
| 2. | Phytomastigophorea | 990 | 1048 | 856 | 2894 |
| | Total | 2392 | 2568 | 1820 | 6780 |

2011-12

| Sr.No. | Flagellates | Monsoon | Winter | Summer | Total |
|--------|--------------------|---------|--------|--------|-------|
| 1. | Zoomastigophorea | 1240 | 1287 | 1046 | 3573 |
| 2. | Phytomastigophorea | 961 | 1192 | 944 | 3097 |
| | Total | 2201 | 2479 | 1990 | 6670 |

4. Result and Discussion

The BBT carafes were incubated for about 20 days at 37°C (1°C) without shaking. During the hatching period, the fixation and abundance of Legionella and eukaryotic networks in the



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planktonic stage and biofilm were noted. To see whether L. pneumophila develops under test circumstances in the investigated water type, control BBTs using cups vaccinated with L. pneumophila and the protozoan host Hartmannellavermiformis were conducted. H. vermiformis was immunised at a concentration of between 3.3 and 4.8 105 cells per litre. Single-jar control tests were completed.

To determine if the growth of native Legionella spp. occurred in the presence or absence of free-living protozoa, several BBT jars were also hatched at 15°C with ordinary water and separated faucet water (3.0-m-pore size and 47-mm-breadth TSTP Isopore layer [Millipore, Molsheim, France]) (Table 1). Additionally, BBT flagons were hatched at 15°C with biomass from the channel beds of groundwater sources A and B to see whether development of native Legionella spp. occurred in the presence of the vaccine and in the absence of the H. vermiformis strain.

4.1. Preparation of inoculum for L. pneumophila and H. vermiformis

Table 1: Characteristics of the water types tested and results of the controls of the biofilm batch test (BBT) incubated at 37°C, where Hv indicates H

| Drigin of sample | Temp | Conc. of ATP | Conc. of Hv | Lp growth | Lp/Hv-ratio |
|--|-------------------|------------------------------------|---------------------------------------|-------------------------|-------------|
| Sign of sample | (°C) ^a | (ng l ⁻¹) ^a | (cells 1 ⁻¹) ^a | in control ^b | control |
| Drinking water supplies | | | | | |
| Groundwater supply A | | | | | |
| Treated water | 11 | <1 | <0.5 | $N.D.^{f}$ | N.A.9 |
| Biomass from filter bed (limestone) | 11 | $4.6 \pm 0.04^{d,e}$ | $<2^e$ | + | 2.1; 2.5 |
| Installation A1 | 39 | 1.9 ± 0.3 | 51 ± 4^d | + | 3.6 |
| Installation A2 | 37 | 3.3+0.2 | 8.2+2.2 | + | 2.9 |
| Groundwater supply B | | | | | |
| Treated water | 11 | 6.7±0.3 | <0.5 | N.D. | N.A |
| Biomass from filter bed (sand) | 11 | 52.6 ± 0.7^{e} | $11.4{\pm}7.6^{e}$ | + | 2.4; 3.5 |
| Installation B1 | 35 | 4.1±0.2 | 6.8 ± 2.5 | + | 2.4 |
| Installation B2 | 37 | 7.8±0.9 | 1530 ± 290 | + | 3.3 |
| Flushed tap water | 12 | 16.0 ± 3.3 | 805±74 | + | 3.0 |
| Groundwater supply C; tap water | 18 | 3.3±0.4 | <4 | + | 3.1 |
| Surface water supply D | | | | | |
| Biomass from filter bed (granular activated carbon) | 5 | 22.1 ± 1.3^{e} | 5.0 ± 1.8^{e} | + | 2.0 |
| Biomass from filter bed (sand) | 5 | 55.5±4.3 ^e | 12.3 ± 1.7^{e} | + | 2.0 |
| Surface water | | | | | |
| Storage reservoir for surface water supply D | 5 | $14.2{\pm}0.5$ | <6.7 | + | 3.5 |
| Water of river Rhine (autumn) | 10 | 83.2±5.4 | <10 | N.D. | N.A |
| Water of river Rhine (winter) | 4 | 61.1±2.4 | <10 | + | 3.0 |
| Treated sewage | 15 | 705.8±4.9 | 28.0 ± 10.8 | + | 1.9 |
| Water from cooling tower | | | | | |
| Cooling tower 1 (pH 7.3) | 27 | $101.8 {\pm} 1.5$ | <6.6 | + | 2.6 |
| Cooling tower 2 (pH 8.5) | 9 | 35.7±0.3 | <2 | + | 3.2 |
| Cooling tower 3 (pH 7.8) | 24 | $19.0 {\pm} 0.1$ | <2 | _ | |
| Cooling tower 4 (pH 6.6) | 19 | 14.7 ± 0.6 | <2 | _ | |



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H. vermiformis was present, but L. pneumophila did not develop in commencing BBT cups that were hatched at 37°C with biomass from supply A's (underlying grouping of 7.9 ng ATP litre) and supply B's (131.7 ng ATP litre) channel beds without L. pneumophila vaccine (Fig. 3.1). Additionally, in the BBT cups with two different types of treated water and no exogenous proliferation of H. vermiformis, vaccine-induced L. pneumophila did not multiply. In contrast, both BBT cups containing water from cooling tower 1 showed critical growth (p 0.025) of L. pneumophila in which H. vermiformis was not identified at day 0 (20 cells liter).

In this water, H. vermiformis also experienced a critical development within a few stretches of brooding, along with the growth of L. pneumophila, which reached its maximum level of development after about 10 days. Based on these observations, L. pneumophila was introduced to all BBT jars to ensure its presence in the experiment, and control cups containing L. pneumophila and H. vermiformis were kept for each test to see whether the water being examined supports protozoan growth in the presence of a host protozoan. In 16 of the 18 control BBT flagons, L. pneumophila showed enormous growth (p 0.025).

5. Conclusion

A variety of geometric shading elements, including the eyelids and brow ridge, as well as behavioural characteristics of vision must be taken into consideration when designing apparatus that is designed to assess the photo biological dosage to ocular tissues. The difficulty of trying to precisely assess the photo biologically important exposure of the cornea, lens, and retina to ultraviolet, visible, and infrared light is complicated by these geometrical and imaging considerations. Actually, only a very minuscule 270 percent of the world's UV irradiance is received by the human eye (diffuse plus direct radiation incident upon a horizontal surface). As a result, the acceptance angle (field of vision) needs to be similar to what the human eye sees. This acceptance angle changes with sky brightness, however. An equipment designed to accurately assess the UVR exposure dosage to the cornea and lens must thus have an adjustable field of view. A mannequin with UV detectors at the ocular locations may be used to assess the UV exposure to the front portion of the eye. Measurements can be done with and without UV-absorbing eyewear or as would be the case when a human wears various kinds of sunglasses. The UV transmittance of the sunglass lenses and the kind of sunglass frame both has a



significant impact on exposure. Certain ocular tissues may experience UV exposures that are equivalent to or greater than those while not wearing sunglasses in various circumstances.

References

- Abou-Akkada, A. R., Howard, B. H. 1961. The biochemistry of rumen protozoa. 4. Decomposition of pectic substances.
- Biochem. J. 78: 512-517. Agrawal, L. K., Harada, H., Tseng, I. C., Okui, H. 1997. Treatment of dilute wastewater in a UASB reactor at a moderate temperature: Microbiological aspects. J. Ferment.
- Bioeng. 83: 185-190. Ajit, H., Suresh, S., Chitra, K. R., Manila[, V. B. 2005. The Buoyant Filter Bioreactor: a high-rate anaerobic reactor for complex wastewaterprocess dynamics with dairy effluent. Water Res. 39: 993 - 904.
- 4. Akin, D. E., Benner, R. E. 1988. Degradation of polysaccharides and lignin by ruminal bacteria and fungi. Appl. Environ. Microbiol. 54: 1117-1125.
- 5. AI-Shahwani, S. M., Horan, N. J. 1991. The use of protozoa to indicate changes in the performance of activated sludge plants. Water Res. 25: 633-638.
- Amann, R. I., Binder, B. J., Olson, R. J., Chisholm, S. W., Devereux, R., Stahl, D. A. 1990a. Combination of 16S rRNA-targeted oligonucleotide probes with flow cytometry for analyzing mixed microbial populations.
- Appl. Environ. Microbiol. 56: 1919-1925. Amann, R. I., Krumholz, L., Stahl, D. A. 1990b. Fluorescent oligonucleotide probes of whole cells for determinative, phylogenetic and environmental studies in microbiology. J.
- Bacteriol. 172: 762-770. Amann, R. I., Springer, N., Ludwig, W., Gartz, H. D., Schleifer, K. H. 1991. Identification in situ and phylogeny of uncultured bacterial endosymbionts. Nature 351: 161-164.
- Ana, M. G., Silvia, D., Sara, B., Andrea, G., Juan, C. G. 2006. Cytotoxicity and bioaccumulation of heavy metals by ciliated protozoa isolated from urban wastewater treatment plants. Res. Microbiol. 15 7:108-118.
- Anderson, G. K., Yang, G. 1992. Determination of bicarbonate and total volatile acid concentration in anaerobic digesters using a simple titration. Water Environ. Res. 64: 53-59. Anja, S., Yvonne, F., Harmut, A. 2005. Detritus as a potential source for



protozoans: utilization of fine particulate plant detritus by a heterotrophic flagellate, Chilomonas paramecium, and, a ciliate, Tetrahymenapyriformis.

- Aqua. Ecol. 39: 439-445. 196 Aoki, K., Tanaka, T., Shinke, R., Nishira, H. 1979. Detection of tannase in polyacrylamide gels. J.
- 12. Chromatogr. 170: 446-448. APHA. AWWA, WEF. 1998. Standard Methods for Examination of Wastes and Wastewater. Washington, DC, USA.
- Arndt, H., Kroeker, M., Nixdorf, B., Kohler, A. 1993. Long-term annual and seasonal changes of meta- and protozooplankton in Lake Mi.iggelsee (Berlin): Effects of eutrophication, grazing activities, and the impact of predation. Int. Rev. Ges. Hydrobiol. 78: 379-402.
- 14. Arora, S. D., Paramjit, K. G. 2001. Comparison of two assay procedures for lignin peroxidase. Enz. Microb. Technol. 28: 602-605.
- Azam, F., Fenchel, T., Field, J. G., Gray, J. S., Meyer-Reil, L. A., Thingstad, F. 1983. The ecological role of water column microbes in the sea. Mar. Ecol. Prog. Ser. 10: 257-263.
- 16. Bakers, S. B., Summerson, W. H. 1941. The colorimetric determination of lactic Acid in biological material. J. Biol. Chem. 138: 535-554.
- 17. Ball, G. H. 1969. Organisms living on and in protozoa. In: Research in Protozoology (Chen, T.T. Ed.). Pergamon Press, London and New York.
- Batstone, D. J., Keller, J., Angelidaki, I., Kalyuzhnyi, S. V., Paviostathis, S. G., Rozzi, A., Sanders, W. T. M., Siegrist, H., Vavilin, V. A. 2002. The IWA Anaerobic digestion model No l(ADMl). Water Sci. Technol. 45: 65-73.
- 19. Battin, T. J., Kaplan, L.A., Newbold, J. D., Hansen, C. M. E. 2003. Contributions of microbial biofilms to ecosystem processes in stream mesocosms. Nature 426: 439-442.
- 20. Beaver, J. R., Crisman, T. L. 1989. The role of ciliated protozoa in pelagic freshwater ecosystems.Microb. Ecol. 17: 111-136

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